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Inferring the flow properties of epithelial tissues from their geometry

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PAPER

Inferring the flow properties of epithelial tissues from their geometry

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E-mail: marko.popovic@epfl.ch and matthieu.wyart@epfl.chKeywords: tissue mechanics, vertex model, plasticity, T_1 transition**Abstract**

Amorphous materials exhibit complex material properties with strongly nonlinear behaviors. Below a yield stress they behave as plastic solids, while they start to yield above a critical stress Σ_c . A key quantity controlling plasticity which is, however, hard to measure is the density $P(x)$ of weak spots, where x is the additional stress required for local plastic failure. In the thermodynamic limit $P(x) \sim x^\theta$ is singular at $x = 0$ in the solid phase below the yield stress Σ_c . This singularity is related to the presence of system spanning avalanches of plastic events. Here we address the question if the density of weak spots and the flow properties of a material can be determined from the geometry of an amorphous structure alone. We show that a vertex model for cell packings in tissues exhibits the phenomenology of plastic amorphous systems. As the yield stress is approached from above, the strain rate vanishes and the avalanches size S and their duration τ diverge. We then show that in general, in materials where the energy functional depends on topology, the value x is proportional to the length L of a bond that vanishes in a plastic event. For this class of models $P(x)$ is therefore readily measurable from geometry alone. Applying this approach to a quantification of the cell packing geometry in the developing wing epithelium of the fruit fly, we find that in this tissue $P(L)$ exhibits a power law with exponents similar to those found numerically for a vertex model in its solid phase. This suggests that this tissue exhibits plasticity and non-linear material properties that emerge from collective cell behaviors and that these material properties govern developmental processes. Our approach based on the relation between topology and energetics suggests a new route to outstanding questions associated with the yielding transition.

1. Introduction

A fascinating aspect of biological systems is their ability to grow into well-defined shapes [1]. Thinking about tissues as materials, what should their properties be to allow for robust morphogenesis? One view is that tissues are viscoelastic fluids, molded into desired shapes by surface tension and active forces [2–9]. An alternative picture is that they are yield stress materials [10] similar to clay. Such materials allow for great control, since shape is changed only if the magnitude of shear stress Σ is above the threshold yield stress Σ_c . These approaches can be thought of as two extremes of a continuous spectrum of models, since at finite temperature, or at finite level of active stress and cell divisions in biological systems [3, 11], materials always eventually flow. Experimental evidence of glassy behavior [12–14] indeed suggests relevance of intermediate cases. Quantitatively, an interesting observable to distinguish these regimes is the ratio between the strain rate increment $\delta\dot{\gamma}$ and the stress increment $\delta\Sigma$ causing it. This ratio is simply $\dot{\gamma}/\Sigma$ for a Newtonian liquid,

but is infinite at Σ_c in a yield stress material at zero temperature. As discussed below, this divergence is associated with collective events where large chunks of the material rearrange. This fact suggests that one may be able to decide in which regime tissues operates simply by imaging their dynamics and geometry. One of our aims is to build the first steps of this long term goal. Note that this endeavor is distinct from non-invasive force inference methods [15, 16], in which one seeks to reconstruct stress—instead of plasticity and rheological properties—from geometry.

As it turns out, there is currently a considerable interest in understanding the relationship between geometry and plasticity in particulate amorphous materials [17–21]. Flow is mediated by local rearrangements termed shear transformations [22] that are coupled by long range elastic interactions [23]. If thermal fluctuations are small, for $\Sigma > \Sigma_c$ the flow consists of avalanches of correlated shear transformations. The characteristic avalanche size diverges at Σ_c [24, 25], and is system spanning for non-stationary slow (quasi-static) flows occurring in the solid phase $\Sigma < \Sigma_c$ [26]. At a macroscopic level, for $\Sigma \geq \Sigma_c$ the shear rate is singular and follows the Herschel–Bulkley law $\dot{\gamma} \sim (\Sigma - \Sigma_c)^\beta$ [27]. A key ingredient of the scaling theory of this phase transition [28] is that the density $P(x)$ of shear transformations at a distance x to their local yield stress (i.e. the shear transformations that yield if the shear stress is increased by x) follows $P(x) \sim x^\theta$, with $\theta > 0$ [29, 30, 31], which is directly related to the presence of extended avalanches [32]. The exponent θ is predicted [33] to vary non-monotonically as shear strain is increased from an isotropic state, as observed in particle-based models [34–36], whereas for $\Sigma > \Sigma_c$ there are no singularities in $P(x)$ and $\theta = 0$ [33]. Various computationally expensive numerical methods are being developed to extract the field of shear transformations and their associated distance to yield stress x from the structure alone [17–21], as it would allow one to study fundamental questions including the possible localization of plastic strain. In this work we present an alternative approach by showing that some models of disordered materials present the same phase transition, in which this extraction is straightforward.

We consider the vertex model of epithelial tissues [37] in its solid phase where it displays a finite elastic modulus [38, 39]. We first show that its yielding transition is similar to particulate amorphous materials: its flow curve is singular with a Herschel–Bulkley exponent $\beta \approx 1.3$, associated with avalanches of plastic events whose size diverges as $S \sim \dot{\gamma}^{-a}$ and last a duration $\tau \sim \dot{\gamma}^{-c}$ where $a \approx 1/4$ and $c \approx 2/3$. In such a model, just like for dry foams, shear transformations are known to correspond to T_1 events [40]. We argue quite generally and test numerically that in models where the energy function depends on the topology, the distance x to the local yield stress follows $x \sim L$ where L is the bond length between two vertices. It implies that $P(x)$ is readily obtainable from the bond length distribution, from which we extract θ . It is found to change non-monotonically under strain with $\theta \approx 0.5$ – 0.6 in isotropic state and with a similar value at large strain at $\Sigma = \Sigma_c$: $\theta \approx 0.56$ – 0.77 . By contrast, θ vanishes in the liquid phase for $\Sigma > \Sigma_c$. Finally, we measure the bond length distribution in fruit fly wing disc and pupal wing epithelia, and find a similar behavior with $\theta \approx 0.7$ – 0.9 . This measurement suggests the existence of collective effects in tissues and raises the possibility that these materials function in a regime of high sensitivity $\delta\dot{\gamma}/\delta\Sigma \gg \dot{\gamma}/\Sigma$.

2. Flow and loading curves of vertex model

We use the standard vertex model of epithelial tissues [37, 41] where the 2D network of polygonal cells is assigned an energy function

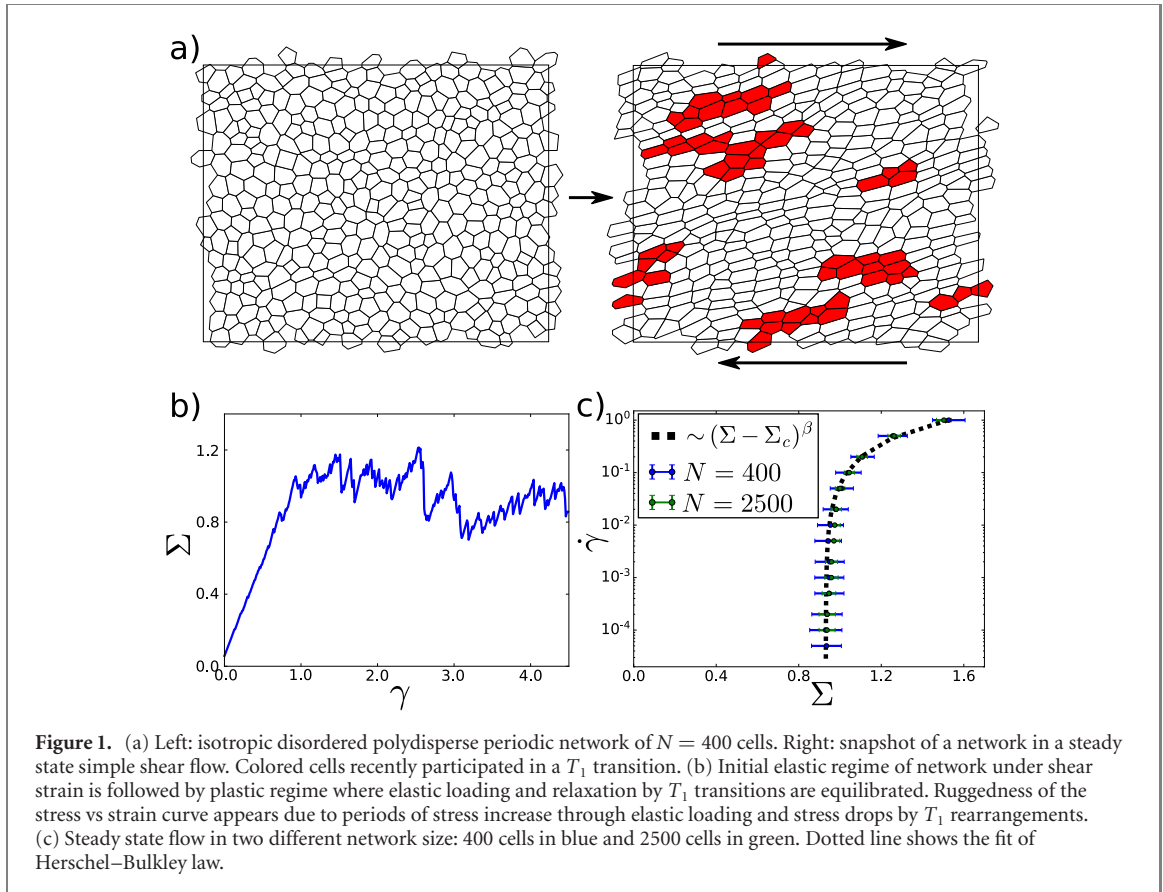
$$E = \sum_{c \in \text{cells}} \frac{1}{2} \left[K(A_c - A_{0,c})^2 + \Gamma_c P_c^2 \right] + \sum_{b \in \text{bonds}} \Lambda_b L_b \quad , \quad (1)$$

where A_c , L_b and P_b are cell area, bond length and cell perimeter, respectively. Model parameters specify preferred cell area $A_{0,c}$, bond tensions Λ_b and cell perimeter stiffness Γ_c . To avoid localisation of flow in a narrow shear band, which occurs in the homogeneous vertex model [42], we introduce cell size polydispersity for shear flows, see appendices A and C. In all simulations the network is in solid phase with the normalised preferred perimeter $p_0 \equiv -\Lambda_b/(2\Gamma_c\sqrt{A_{0,c}}) \simeq 3.41$ for all cells, well below the rigidity transition point $p_0^* \simeq 3.81$ [38]. Note that our results below may not hold in the fluid phase of the vertex model [43], or in active tension networks with isogonal soft modes [44].

The dynamics of the cellular network is described by overdamped dynamics of vertex positions \vec{r}_α

$$\frac{d\vec{r}_\alpha}{dt} = -\nu \vec{\nabla}_\alpha E, \quad (2)$$

where $\nu = 1$ is a mobility. A T_1 transition occurs when a bond length becomes smaller than a threshold length ϵ_{T_1} , see appendix B.



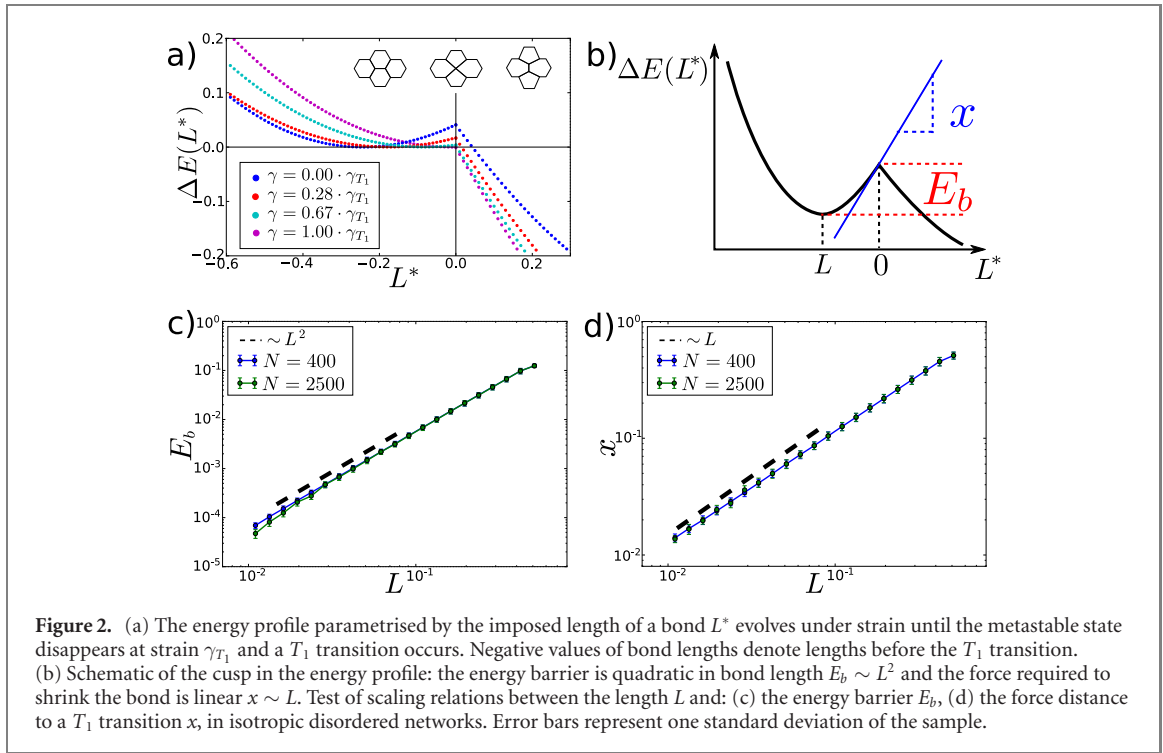
We use two ensembles of isotropic disordered networks with $N = 400$ and $N = 2500$ cells, as described in appendix D. An example of a network with $N = 400$ cells is shown in figure 1(a) left. We perform simple shear strain simulations on these networks at a constant strain rate, illustrated in figure 1(a) right for $\dot{\gamma} = 10^{-4}$. The network initially responds elastically: the shear stress Σ in the network grows almost linearly (figure 1(b)). As the strain is increased, T_1 transitions occur and relax the stress in the network, visible as sudden drops in the stress vs strain curve. In figure 1(a) right we visualise recent T_1 transitions that occurred during a strain increment $\Delta\gamma = 0.15$ by coloring participating cells in red. T_1 transitions appear to be correlated and organised into avalanches of various sizes, corresponding to widely distributed stress drops in figure 1(c). Eventually a steady state is reached in which stress relaxation due to T_1 transitions balances the elastic loading. In figure 1(d) we show the steady state flow curve. It is well described by the Herschel–Bulkley law $\dot{\gamma} \sim (\Sigma - \Sigma_c)^\beta$ [27] with the yield stress $\Sigma_c \approx 0.93$ ⁷ and exponent $\beta \approx 1.3$ ⁸.

3. Energy cusp at T_1 transitions

Next we characterize the T_1 transitions or elementary plastic events. For this purpose, we identify the bond in a network that will first disappear under strain. We then constrain the length of that bond to a value L^* and determine the energy of the network under strain. The original bond length is assigned negative values and the new bond that appears through the T_1 transition is assigned positive values. In figure 2 we show the energy of the network relative to the energy of the unconstrained network, as a function of strain $\Delta E(L^*; \gamma) \equiv E(L^*; \gamma) - E(\gamma)$. Originally the system is in a metastable state, corresponding to the local minimum of ΔE . As the shear stress increases the minimum disappears and the T_1 transition occurs. The energy profile shows a cusp at the onset of T_1 where $L^* = 0$, a well known feature of the vertex model energy landscape [38, 46–48]. The presence of a cusp in the energy profile allows us to relate the bond

⁷ The stress component corresponding to the simple shear is defined by $\Sigma \equiv (\partial E / \partial \gamma) / N$. In our simulations $\Gamma = 1$ and a typical bond length is $L_0 = 1$. Therefore, the reported values of stress can be understood as normalised by ΓL_0^2 .

⁸ A precise measurement of the exponent β that could discriminate theoretical predictions [45] is beyond the scope of this work.



length of short bonds to the additional force⁹ x at the bond needed to drive a T_1 transition. Namely, expanding $\Delta E(L^*, \gamma)$ in bond length around the equilibrium value L reads:

$$\Delta E(L^*) \approx \Delta E(L) + \frac{1}{2} \Delta E''(L) (L^* - L)^2 \quad (3)$$

and we see that at T_1 transition, corresponding to $L^* = 0$, the energy barrier to the T_1 transition is:

$$E_b \equiv \Delta E(L^* = 0) \approx \frac{1}{2} \Delta E''(L) L^2. \quad (4)$$

Thus, the force on that bond required to trigger the T_1 transition is:

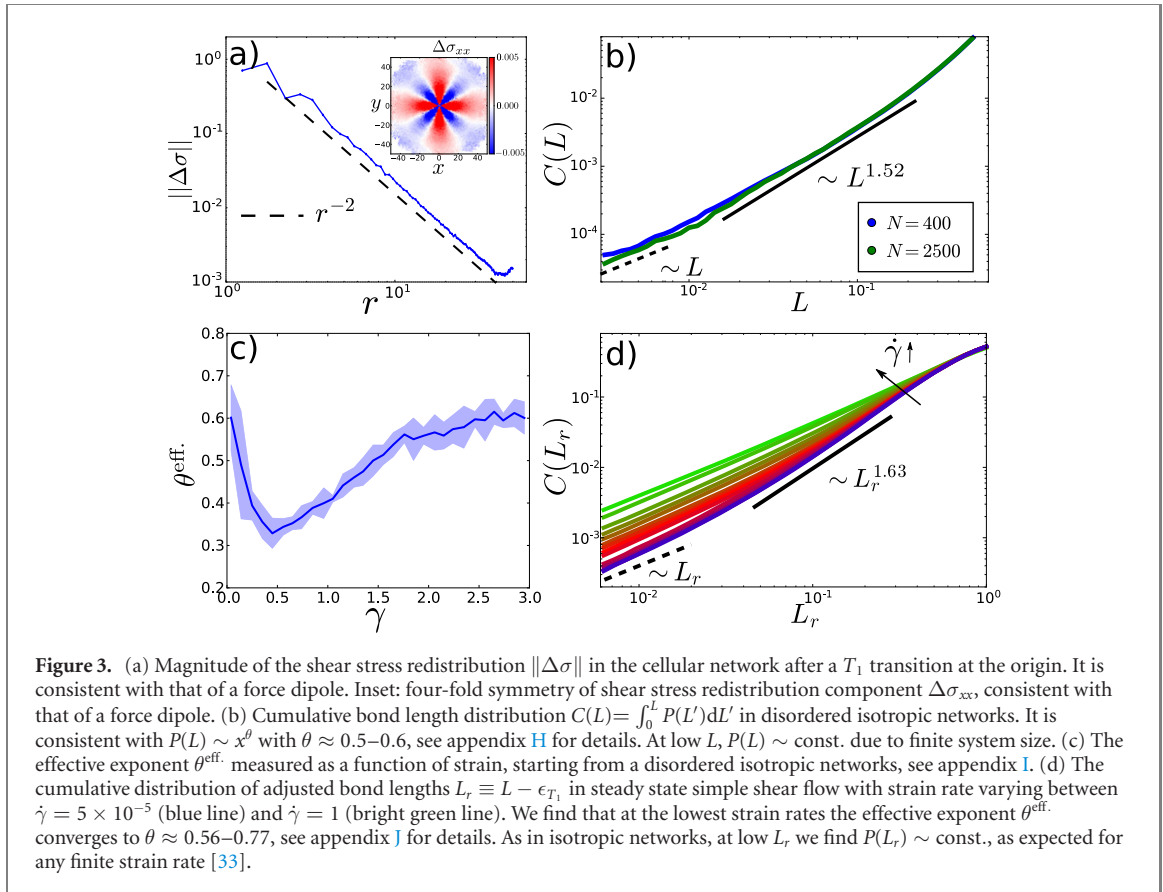
$$x \approx \Delta E''(L) L. \quad (5)$$

Since the effective stiffness of the bond $\Delta E''(L)$ is expected to be finite at a T_1 transition, we find $E_b \sim L^2$ and $x \sim L$. These relationships were previously reported and supported analytically in a continuum elastic model of a T_1 transition [48]. Here, we find that they are a generic consequence of the cusp in the energy profile¹⁰. Ultimately, the origin of this cusp lies in the form of the energy function that depends on the bond lengths, cell perimeters and area. These quantities are smooth functions of the vertex positions and the energy function is therefore continuous even when topology changes. However, at a T_1 transition a bond is removed and another one added. This implies that gradients of the energy function are discontinuous at the T_1 transition, leading to a cusp in the energy profile. Consequently, forces can change discontinuously at the transition point. Therefore, we expect to generically find $x \sim L$ in cellular systems such as epithelial tissues and dry foams for which the dependence of the energy on the vertex position is topology-dependent. By contrast, particle systems in which the energy depends on the particle positions independently of any notion of topology cannot show such a cusp (as long as the interaction potential is smooth). Furthermore, due to the cusp at the T_1 transition the stiffness of the corresponding displacement mode does not vanish, as it would at the plastic event in particle systems¹¹. Therefore, we do not expect to

⁹ Note that exerting a stress increment at the boundary of the system will in general generate a bond force proportional to that increment, so the quantity x we use here characterizes well the distance to a local yield stress.

¹⁰ For example, if the energy landscape of vertex model were smooth at T_1 transitions the relationship would be $x \sim \sqrt{L}$ and $E_b \sim L^3$.

¹¹ In systems with smooth energy function a plastic event corresponds to the usual saddle-node bifurcation.



find a signature of local plastic events in the eigenvalues of Hessian matrix of energy function, which could explain the lack of soft non-localised modes recently observed in the Voronoi vertex model¹² [49].

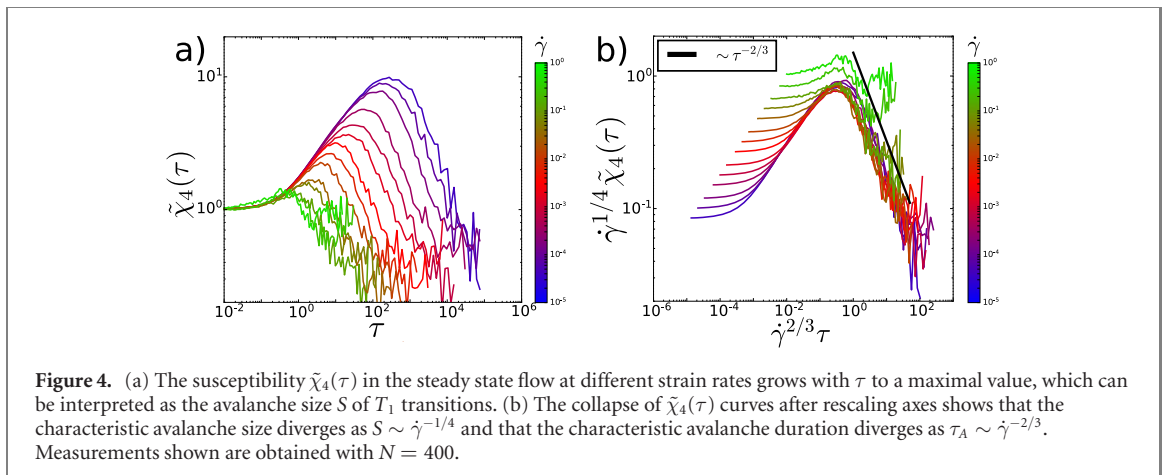
We test these predictions in isotropic disordered networks by forcing bond length to attain a very small value $L_{\min} = 10^{-6}$ and determining the corresponding network energy change and the constraining force magnitude x . Results shown in figures 2(c) and (d) are consistent with our predictions. Therefore, identifying the locations of short bonds allows us to read the map of ‘weak spots’ in the system, as well as to deduce the distribution $P(x)$.

4. Stability of the cellular network

After a T_1 transition, the network relaxes to a new metastable state with redistributed shear stresses. We measure the stress redistribution by enforcing a T_1 transition and measuring the change in stress within each cell after the network had relaxed (the cellular stress is defined as in [50]). In an elastic 2D medium we expect the shear stress redistribution to be consistent with that of a force dipole in an elastic medium: $\Delta\sigma_{xx} \sim \cos(4\varphi)/r^2$, $\Delta\sigma_{xy} \sim \sin(4\varphi)/r^2$ [23]. Figure 3(a) shows the shear stress redistribution, obtained by orienting the disappearing bond direction along the x -axis and averaging over 50 realisations, as detailed in appendix E. We find a clear four-fold symmetry of $\Delta\sigma_{xx}$ (inset) as well as inverse quadratic decay of its magnitude, as expected for a force dipole and consistent with simulations of 2D foams [40].

The stress change after a T_1 transition can trigger new T_1 transitions if there are bonds with small x in the network. In the solid phase, the stability of the network with respect to extensive avalanches of T_1 transition imposes $P(x) \sim x^\theta$ with $\theta > 0$, otherwise never-ending avalanches would occur [31]. As we have demonstrated $x \sim L$ in the vertex model. Therefore, bond length distribution should vanish with the same exponent $P(L) \sim L^\theta$, and θ can be extracted from $P(L)$. We measure the cumulative distribution $C(L) \equiv \int_0^L P(L')dL'$ in disordered isotropic networks. We find a scaling regime, whose range of validity grows with system size, for which $\theta \approx 0.5-0.6$ (figure 3(b)), see appendix H for details. At even smaller L ,

¹² The voronoi vertex model has the same energy function as the usual vertex model, but its degrees of freedom correspond to cell centers. The network topology is constructed from these centers by performing a Voronoi tessellation at each time-point.



the bond lengths distribution departs from this scaling as $P(L) \sim \text{const.}$, as expected due to finite size effects and also observed in elasto-plastic models [28].

Interestingly, these values are consistent with those found in two-dimensional elasto-plastic models [28]. In these coarse-grained models, the material is described as a collection of mesoscopic blocks with a simplified description of plastic events: a block yields when the local yield stress is reached, it accumulates plastic strain and redistributes stress in the material as a force dipole [51, 52]. Since both ingredients are present in vertex model as well, as we have seen, it is not surprising that we find a consistent value of θ .

We next studied the evolution of the exponent θ during the transient loading period, between the initially isotropic network and the steady state. Surprisingly, it has been predicted that θ would then non-monotonically depend on strain [33], a result observed in elasto-plastic models [26] but only indirectly observable in amorphous solids where $P(x)$ is very hard to access [34–36]. To test directly this prediction, we measure the bond length distribution as a function of strain at a small constant strain rate $\dot{\gamma} = 10^{-4}$, close to the quasi-static limit. Note that even in the thermodynamic limit we expect to find singular $P(x)$ and $P(L)$ only in the quasi-static limit of vanishing strain rate (at any finite rate, there are always T_1 transition occurring leading to $\theta = 0$ [33]). However, in a finite system we can still measure an effective exponent θ^{eff} . We confirm that the evolution of θ^{eff} with strain is non-monotonic, see figure 3(c).

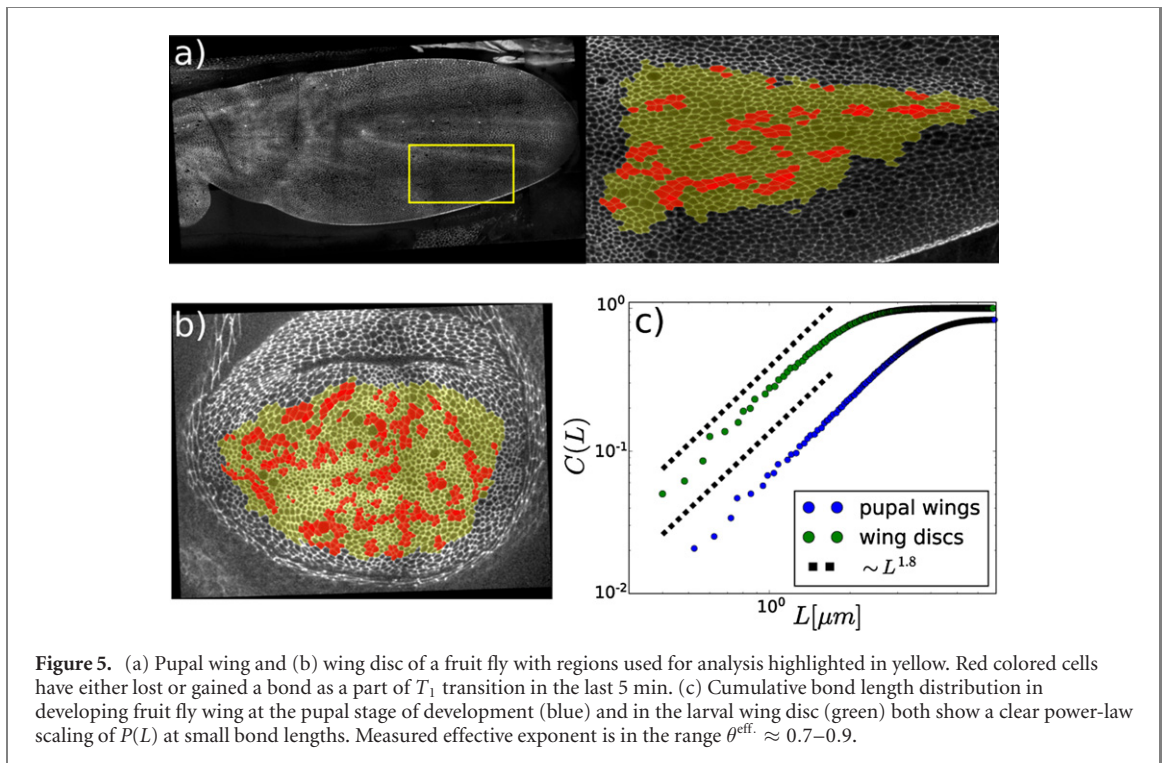
It is also important to quantify the distribution of bond lengths in steady state flow, see figure 3(d). At high strain rates we find $\theta^{\text{eff}} \rightarrow 0$ as expected, while in the limit of vanishing strain rates the effective exponent approaches the value $\theta \approx 0.56\text{--}0.77$ (figure 3(d)). Thus, $P(L)$ can be used to locate the distance to the yield stress, at least in this setting where noise is absent.

Finally, note that the fact that the density of bonds vanishes at small argument as $P(L) \sim L^\theta$ is a necessary constraint required by network stability [31]. Such a power-law distribution of weak regions is found in amorphous materials under athermal quasi-static shear and in slowly driven glassy materials in various contexts [32]. Generating glasses in which weak regions are depleted even further requires extreme annealing [53]. Cellular networks could also be made more stable, for example by sequentially performing T_1 transitions on the shortest bonds as discussed in reference [54]. Such an annealing process depletes the distribution of short bonds and can create a gap in the distribution $P(L)$ that does not occur in the distributions discussed here. More generally, we expect our results to be relevant for describing disordered developing tissues undergoing shape changes through T_1 transitions, whereas it might not describe very well-annealed tissues.

5. Collective behavior of T_1 transitions

Quite generally in disordered systems [31], a singularity in the density of weak regions $P(x)$ is synonymous to avalanche-type response where many weak regions—here T_1 's—rearrange in concert. To test if this idea holds in the vertex model, we quantify the correlation between T_1 transitions by using a susceptibility motivated by the four-point susceptibility χ_4 studied in glasses [55, 56], elasto-plastic models [57, 58] and vertex models [49]. We define $\tilde{\chi}_4$ as a normalised variance of the number $n_{T_1}(\tau)$ of T_1 transitions in the time-window τ [57, 59]:

$$\tilde{\chi}_4(\tau) \equiv \frac{\langle (n_{T_1}(\tau) - \langle n_{T_1}(\tau) \rangle)^2 \rangle}{\langle n_{T_1}(\tau) \rangle} \quad (6)$$



If T_1 transitions were completely independent, the variance would be equal to the mean at any τ and $\tilde{\chi}_4(\tau) = 1$. For T_1 transitions organised in avalanches $\tilde{\chi}_4$ grows and reaches a maximum at τ_A , corresponding to the typical avalanche duration. The value at the maximum can be interpreted as a characteristic avalanche size $S \equiv \tilde{\chi}_4(\tau_A)$. After an avalanche, the stress has relaxed locally and new avalanches are less likely to occur. This effect leads to a decay of $\tilde{\chi}_4(\tau)$ for $\tau > \tau_A$.

We measure $\tilde{\chi}_4(\tau)$ in steady state shear flow at different strain rates, see figures 4(a) and (b). We find that $\tilde{\chi}_4$ displays a peak which grows near the transition point $\dot{\gamma} \rightarrow 0$, indeed supporting the idea that the dynamics becomes collective at that point. Furthermore, we find that the $\tilde{\chi}_4$ curves at different strain rates collapse when re-scaling the axes as $\dot{\gamma}^{2/3}\tau$ and $\dot{\gamma}^{1/4}\tilde{\chi}_4$. Therefore, as the strain rate vanishes, the mean avalanche size diverges as $S \sim \dot{\gamma}^{-1/4}$ and the mean avalanche duration as $\tau_A \sim \dot{\gamma}^{-2/3}$.

6. Fly wing epithelia

We have shown that in the vertex model, the bond length distribution $P(L)$ is indicative of the regime in which the material flows: it presents a singular distribution approaching the solid phase, where the dynamics becomes collective and the flow curve is non-linear.

As a first test of the relevance of these ideas to real tissues, we analyze the bond length distribution in wing epithelium of the fruit fly at two stages of development: (i) during pupal wing morphogenesis, imaged *in vivo* [6] and (ii) the wing disc epithelium, in third instar larva wing disc epithelium imaged *ex vivo* [60]. In the pupal wing we considered a region defined by the longitudinal veins denoted L4 and L5, and the posterior crossvein (yellow cells in figure 5(a)), imaged at 5 min intervals between 19 and 23 h after puparium formation, collected from 3 experiments [61]. In figure 5(b) we show the analyzed region in the wing disc epithelium, corresponding to the wing disc pouch, imaged at 5 min intervals over about 13 h and collected from 5 experiments. In both figures 5(a) and (b) we indicate in red the cells that have lost or gained a bond as a part of T_1 transition in the last 5 min (the time resolution of experiments).

Note that the wing disc epithelia have been developing for about 100 h before the imaging has started [60], while pupal wings have undergone a significant three-dimensional shape change, called eversion [62], before the pupal morphogenesis. Therefore, the initial state of these tissues at the beginning of the experiments could already contain a significant strain history. Thus, although the amount of strain accumulated during experiments is small ($\sim 0.1-0.2$ for pupal wings [6, 61] and $\sim 0.05-0.1$ for wing discs [60]), it is not clear if we are in a large or small strain regime, with respect to the strain where figure 3(c) displays a minimum.

Remarkably, we find that in both tissues the bond length distribution $P(L)$ vanishes at small L with the effective exponent $\theta^{\text{eff.}} \approx 0.7-0.9$ (see figure 5(c) and appendix K for details), similar to those measured in

the slowly flowing vertex model at small or large strain. This observation suggests that the scaling relation $x \sim L$ holds in real tissues as well, in the range of L that we can probe. Clearly for very small bonds, this relationship must eventually break down due to finite size of vertices [63, 64].

It would be very interesting to test this scaling relation directly by perturbing the system. It could be achieved by observing tissue response to a localised mechanical perturbation, such as laser ablation of a single bond: if the energy landscape were smooth with no cusps, the stiffness of the corresponding displacement mode would vanish in approach to a T_1 transition as expected near a saddle node bifurcation. As a consequence, a strong locally heterogeneous response would be observed in the experiment at locations of short bonds just before they rearrange, as observed in particulate amorphous solids [65] preceding a plastic event. On the other hand, in presence of a cusp there would be no softening and no strong locally heterogeneous displacement preceding cell rearrangements.

Our observation also raises the possibility that developing tissues can lie in a non-linear regime with $\delta\dot{\gamma}/\delta\Sigma \gg \dot{\gamma}/\Sigma$, where collective effects are important. Unfortunately, such collective effects are very hard to measure in our experimental data, because the strain rate is not stationary, leading to difficulties using the definition of $\tilde{\chi}_4$. Thus, it would be important to look for non-linear effects more directly by measuring stress dynamics using laser ablation experiments and comparing them to elastic and plastic flow components [6, 66–68]. Alternatively, epithelia obtained as cultured cell monolayers might provide interesting experimental systems that allow for direct rheological experiments. In such systems non-linear flow properties have been observed [69] and bond length distributions $P(L)$ could be measured in different flow regimes.

7. Discussion

We have shown that in the vertex model in its solid phase, the distribution of bond lengths provides the distribution of local distances to yield stress x . The distribution $P(x)$ reveals properties of the regime in which flow is occurring. This result has two consequences.

First in developing epithelia, we observe a singular distribution of bond lengths, consistent with the one found in the vertex model. This result raises the intriguing possibility that non-linear collective effects may be important in tissues, and suggests further empirical tests. Yet, active forces and cell divisions or extrusions could affect the relationship between geometry and flow properties at shear rates comparable or smaller than cell division and extrusion rates [3, 11]. Extending our results to this flow regime will require incorporating these active processes. From a theoretical perspective, how the density of weak spots depends on stress in the presence of noise—even a simple thermal noise—is not well understood in amorphous solids, and is just starting to be investigated [11, 70]. In this light, it would be important to study in the future how different kinds of noise affect the flow curve and the distribution $P(L)$ in the vertex model.

Secondly, despite the fact that the energy functional of the vertex model is more complex than that of usual particulate materials (in which interaction can be radially symmetric), the relationship between geometry and the presence of weak spots is much simpler in the vertex model. Outstanding questions in the context of amorphous materials, such as predicting how amorphous solids break by forming shear bands in which most plastic events occur, are hampered by the difficulty of measuring the distribution of weak spot with small x [17–21]. The vertex model may thus be ideal to understand the universal aspects by which amorphous materials break and flow.

Acknowledgments

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Appendix A. Vertex model parameters

We used the following parameter values in most simulations:

- $K_c = 10$
- $\Lambda_b = -11$

- $A_{0,c}$ and Γ_c were always chosen so that $p_0 = -\Lambda_b/(2\Gamma_c\sqrt{A_{0,c}}) = 11/(2\sqrt{3\sqrt{3}/2}) \approx 3.41$ is constant throughout the network.
 - $\nu = 1$
- Any change of parameters in a particular simulation is explicitly listed below.

Appendix B. T_1 transition implementation

When a bond length becomes smaller than a threshold value ϵ_{T_1} a T_1 transition is attempted: old bond and corresponding vertices are destroyed and new ones are created, then forces on the new bond are computed and T_1 transition is allowed if the tension in the new bond is positive (forces are stretching the new). Otherwise, the T_1 transition is canceled and the network is reverted to the original state. The choice of ϵ_{T_1} is specified for particular simulations below. To avoid the possibility of an extrusion we do not allow bond loss by T_1 transition for cells with 3 neighbors.

Appendix C. Cell size polydispersity

In the flow simulations we avoided crystallization and shear banding by introducing cell size polydispersity: preferred cell areas $A_{0,c}$ are uniformly distributed on the interval $[\bar{A}_{0,c}/2, 3\bar{A}_{0,c}/2]$, where the mean preferred cell area $\bar{A}_{0,c} = 3\sqrt{3}/2$ corresponds to size of regular hexagons before initial network randomization. Parameter Γ_c was then chosen so that all cells have the same value of p_0 . Finally, we set $K_c = 40$.

Appendix D. Isotropic disordered networks

To create isotropic disordered networks, we first create a hexagonal network with bond length $l = 1$. The energy function parameters are then set so that the network is in the solid phase $p_0 \approx 3.7$, but close to the transition point $p_0^* = 3.81$ [38]. The parameters used are:

- $K_c = 1$
- $\Lambda_b = -12$
- $\Delta t = 5 \times 10^{-4}$
- $\epsilon_{T_1} = 5 \times 10^{-4}$

For the randomization process we introduce fluctuations of the bond tension Λ independently in each bond by simulating its dynamics as a time-discretised Ornstein–Uhlenbeck process:

$$\Lambda(t + \delta t) = \Lambda(t) - k(\Lambda(t) - \Lambda_0) + \xi\sqrt{\delta t}, \quad (\text{D1})$$

where $\Lambda_0 = -12$, $\delta t = 10^{-2}$, $k = 1$ and ξ a random variable taken from a normal distribution $\mathcal{N}(0, 1)$.

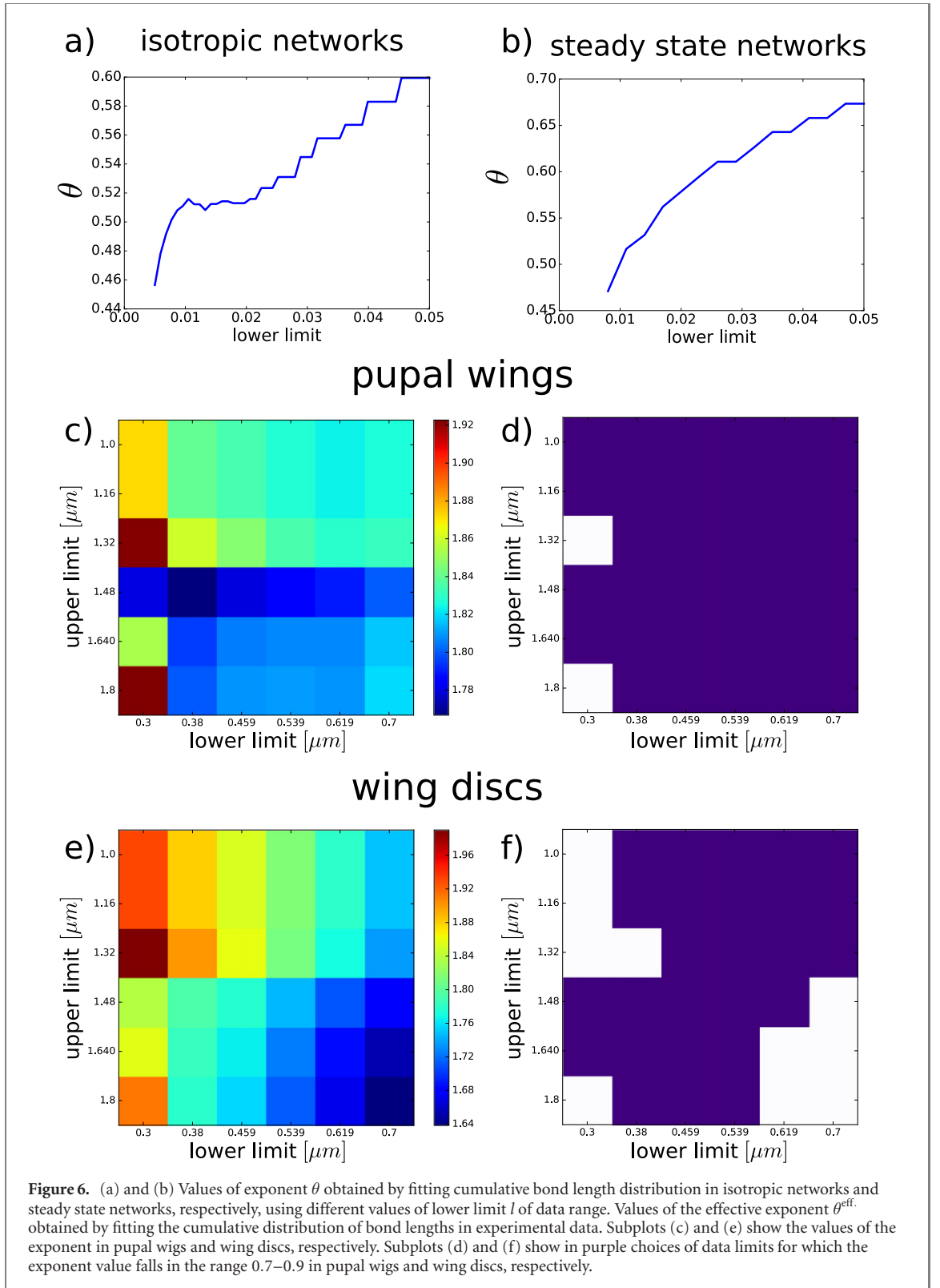
Networks are evolved for $t_r = 50$, then fluctuations are frozen and the network is relaxed for time 0.5. Finally, network parameters are set to simulation values and networks are further relaxed over time using $\epsilon_{T_1} = 10^{-4}$ until the net force on any vertex was below $\epsilon_F = 10^{-4}$.

Appendix E. Stress redistribution by a T_1 transition

Results of the stress redistribution from a T_1 event were obtained from an $N = 2500$ network. After initial relaxation (until net force on any vertex is below $\epsilon_F = 10^{-5}$) 50 bonds of length $l < 0.1$ were selected randomly. Each of these bonds was shrunk below the T_1 threshold. If the tension in the newly formed bond was negative (so that T_1 transition would revert back, see appendix A) the bond was not considered, otherwise the T_1 was performed and the network was relaxed (with $\epsilon_F = 10^{-4}$) and shear stress change for each cell was recorded. We defined cell position as the position of its center of mass and the position was recorded in a coordinate system centered at the middle of the original bond that was selected and whose x -axis was aligned with the original bond direction. Finally, we obtained the average shear stress change in space by performing a cell area-weighted average in spatial bins.

Appendix F. Direct measurement of $P(x)$

In a subset of isotropic disordered networks each bond of length $L < 0.5$ was selected and constrained to be of length $L_c = 10^{-6}$. With this constraint on the bond, the network is then relaxed (until the net force on



any vertex was below $\epsilon_F = 10^{-3}$). Once the relaxation is over, the magnitude of the force acting on the vertices of the constrained bond is recorded as x , as well as network energy change ΔE .

Appendix G. Steady state shear flow

We apply simple shear strain to a network at each time-step using an affine transformation of all vertex positions:

$$\Delta x_\alpha = \frac{y_\alpha}{L_y} \Delta \gamma, \quad (\text{G1})$$

where (x_α, y_α) are coordinates of vertex α , $\Delta\gamma$ is strain increment, and L_y simulation box size in y direction. The applied strain rate $\dot{\gamma} = \Delta\gamma/\Delta t$ was always constant during a simulation. T_1 transition threshold was $\epsilon_{T_1} = 10^{-2}$ and simulation time step $\Delta t = 10^{-3}$.

Appendix H. Isotropic θ

We determined the range of values corresponding to this exponent by fitting cumulative bond length distribution obtained from 50 isotropic networks of size $N = 2500$, which exhibit a broader range of scaling than $N = 400$ networks. We performed the power law fit on a range of data $[l, 0.2]$ with varying lower limit l as shown in figure 6(a). We find that for values of lower limit l in the range $[0.01, 0.05]$ the exponent θ varies between 0.5 and 0.60.

Appendix I. Transient θ_L^{eff}

We fitted the effective exponent on cumulative distribution of adjusted bond lengths $L_r = L - \epsilon_{T_1}$ in the range $[0.03, 0.3]$, accumulated from 700 realisations obtained in $N = 400$ networks at strain rate $\dot{\gamma} = 10^{-4}$, recorded at strain resolution $\delta\gamma_1 = 0.02$. The plot in figure 3(c) was obtained by averaging the results in windows of width $\delta\gamma_2 = 0.1$ with the shaded regions indicating the corresponding standard deviation in each window.

Appendix J. Steady state θ

Cumulative bond length distribution of adjusted bond lengths $L_r = L - \epsilon_{T_1}$ shown in figure 3(d) of the main text are obtained from 1400 networks at strain beyond 5 taken at strain intervals $\Delta\gamma = 0.02$. For the lowest strain rate $\dot{\gamma} = 5 \times 10^{-5}$ we fitted a power law on a range of data $[l, 0.3]$ with varying lower limit l as shown in figure 6(b). We find that for values of lower limit l in the range $[0.02, 0.05]$ the exponent θ varies between 0.56 and 0.77.

Appendix K. Effective exponent in experimental data

To determine the effective exponent θ_L^{eff} in experimental data we fitted a power law to the cumulative distribution of bond lengths. We vary the fitting range to estimate confidence interval of the fits and we find that in most cases fitted exponents fall in the range $0.7 < \theta_L^{\text{eff}} < 0.9$, as shown in figures 6(c)–(f).

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References

- [1] Thompson D W 1945 *On Growth and Form* (Cambridge: Cambridge University Press)
- [2] Bittig T, Wartlick O, Kicheva A, González-Gaitán M and Jülicher F 2008 Dynamics of anisotropic tissue growth *New J. Phys.* **10** 063001
- [3] Ranft J, Basan M, Elgeti J, Joanny J-F, Prost J and Jülicher F 2010 Fluidization of tissues by cell division and apoptosis *Proc. Natl Acad. Sci.* **107** 20863–8
- [4] Lee P and Wolgemuth C W 2011 Crawling cells can close wounds without purse strings or signaling *PLoS Comput. Biol.* **7** 1–8
- [5] Blanch-Mercader C, Casademunt J and Joanny J F 2014 Morphology and growth of polarized tissues *Eur. Phys. J. E* **37** 41
- [6] Etournay R *et al* 2015 Interplay of cell dynamics and epithelial tension during morphogenesis of the drosophila pupal wing *eLife* **4** e07090
- [7] Banerjee S, Kazage J, Utuje C and Cristina Marchetti M 2015 Propagating stress waves during epithelial expansion *Phys. Rev. Lett.* **114** 228101
- [8] Popović M, Nandi A, Merkel M, Etournay R, Eaton S, Frank J and Salbreux G 2017 Active dynamics of tissue shear flow *New J. Phys.* **19** 033006
- [9] Frank J, Grill S W and Salbreux G 2018 Hydrodynamic theory of active matter *Rep. Prog. Phys.* **81** 076601
- [10] Mongera A *et al* 2018 A fluid-to-solid jamming transition underlies vertebrate body axis elongation *Nature* **561** 401–5
- [11] Matoz-Fernandez D A, Agoritsas E, Barrat J-L, Bertin E and Martens K 2017 Nonlinear rheology in a model biological tissue *Phys. Rev. Lett.* **118** 158105

- [12] Angelini T E, Hannezo E, Trepant X, Marquez M, Fredberg J J and Weitz D A 2011 Glass-like dynamics of collective cell migration *Proc. Natl Acad. Sci.* **108** 4714–9
- [13] Nnetu K D, Knorr M, Käs J and Zink M 2012 The impact of jamming on boundaries of collectively moving weak-interacting cells *New J. Phys.* **14** 115012
- [14] Schötz E-M, Lanio M, Talbot J A and Manning M L 2013 Glassy dynamics in three-dimensional embryonic tissues *J. R. Soc. Interface* **10** 20130726
- [15] Ishihara S and Sugimura K 2012 Bayesian inference of force dynamics during morphogenesis *J. Theor. Biol.* **313** 201–11
- [16] Chiou K K, Hufnagel L and Shraiman B I 2012 Mechanical stress inference for two dimensional cell arrays *PLoS Comput. Biol.* **8** e1002512
- [17] Cubuk E D, Schoenholz S S, Rieser J M, Malone B D, Rottler J, Durian D J, Kaxiras E and Liu A J 2015 Identifying structural flow defects in disordered solids using machine-learning methods *Phys. Rev. Lett.* **114** 108001
- [18] Gartner L and Lerner E 2016 Nonlinear modes disentangle glassy and Goldstone modes in structural glasses *SciPost Phys.* **1** 016
- [19] Patinet S, Vandembroucq D and Falk M L 2016 Connecting local yield stresses with plastic activity in amorphous solids *Phys. Rev. Lett.* **117** 045501
- [20] Wijtmans S and Manning M L 2017 Disentangling defects and sound modes in disordered solids *Soft Matter* **13** 5649–55
- [21] Schwartzman-Nowik Z, Lerner E and Bouchbinder E 2019 Anisotropic structural predictor in glassy materials *Phys. Rev. E* **99** 060601
- [22] Argon A S 1979 Plastic deformation in metallic glasses *Acta Metall.* **27** 47–58
- [23] Picard G, Ajdari A, Lequeux F and Bocquet L 2004 Elastic consequences of a single plastic event: a step towards the microscopic modeling of the flow of yield stress fluids *Eur. Phys. J. E* **15** 371–81
- [24] Lemaître A and Caroli C 2009 Rate-dependent avalanche size in athermally sheared amorphous solids *Phys. Rev. Lett.* **103** 065501
- [25] Nicolas A, Ferrero E E, Martens K and Barrat J-L 2018 Deformation and flow of amorphous solids: insights from elastoplastic models *Rev. Mod. Phys.* **90** 045006
- [26] Lin J, Gueudré T, Rosso A and Wyart M 2015 Criticality in the approach to failure in amorphous solids *Phys. Rev. Lett.* **115** 168001
- [27] Herschel W H and Bulkley R 1926 Konsistenzmessungen von Gummi-Benzollösungen *Kolloid-Z.* **39** 291–300
- [28] Lin J, Lerner E, Rosso A and Wyart M 2014 Scaling description of the yielding transition in soft amorphous solids at zero temperature *Proc. Natl Acad. Sci. USA* **111** 14382–7
- [29] Lemaître A and Caroli C 2007 Plastic response of a 2d amorphous solid to quasi-static shear: II-dynamical noise and avalanches in a mean field model (arXiv:0705.3122)
- [30] Karmakar S, Lerner E and Procaccia I 2010 Statistical physics of the yielding transition in amorphous solids *Phys. Rev. E* **82** 055103
- [31] Lin J, Saade A, Lerner E, Rosso A and Matthieu Wyart M 2014 On the density of shear transformations in amorphous solids *Europhys. Lett.* **105** 26003
- [32] Müller M and Wyart M 2015 Marginal stability in structural, spin, and electron glasses *Annu. Rev. Condens. Matter Phys.* **6** 177–200
- [33] Lin J and Wyart M 2016 Mean-field description of plastic flow in amorphous solids *Phys. Rev. X* **6** 011005
- [34] Ji W, Popović M, de Geus T W J, Lerner E and Wyart M 2019 Theory for the density of interacting quasilocated modes in amorphous solids *Phys. Rev. E* **99** 023003
- [35] Ozawa M, Berthier L, Biroli G, Rosso A and Tarjus G 2018 Random critical point separates brittle and ductile yielding transitions in amorphous materials *Proc. Natl Acad. Sci. USA* **115** 6656–61
- [36] Shang B, Guan P and Barrat J-L 2019 Elastic avalanches reveal marginal behaviour in amorphous solids (arXiv:1908.08820)
- [37] Farhadifar R, Röper J-C, Aigouy B, Eaton S and Jülicher F 2007 The influence of cell mechanics, cell–cell interactions, and proliferation on epithelial packing *Curr. Biol.* **17** 2095–104
- [38] Bi D, Lopez J H, Schwarz J M and Manning M L 2015 A density-independent rigidity transition in biological tissues *Nat. Phys.* **11** 1074–9
- [39] Bi D, Yang X, Cristina Marchetti M and Lisa Manning M 2016 Motility-driven glass and jamming transitions in biological tissues *Phys. Rev. X* **6** 021011
- [40] Kabla A and Debrégeas G 2003 Local stress relaxation and shear banding in a dry foam under shear *Phys. Rev. Lett.* **90** 258303
- [41] Staple D B, Farhadifar R, Röper J-C, Aigouy B, Eaton S and Jülicher F 2010 Mechanics and remodelling of cell packings in epithelia *Eur. Phys. J. E* **33** 117–27
- [42] Merkel M 2014 From cells to tissues: remodeling and polarity reorientation in epithelial tissues *PhD Thesis* Technische Universität Dresden
- [43] Park J-A *et al* 2015 Unjamming and cell shape in the asthmatic airway epithelium *Nat. Mater.* **14** 1040–8
- [44] Noll N, Mani M, Heemskerk I, Streichan S J and Shraiman B I 2017 Active tension network model suggests an exotic mechanical state realized in epithelial tissues *Nat. Phys.* **13** 1221–6
- [45] Ferrero E E and Jagla E A 2019 Criticality in elastoplastic models of amorphous solids with stress-dependent yielding rates *Soft Matter* **15** 9041–55
- [46] Bi D, Lopez J H, Schwarz J M and Manning M L 2014 Energy barriers and cell migration in densely packed tissues *Soft Matter* **10** 1885
- [47] Su T and Lan G 2016 Overcrowding drives the unjamming transition of gap-free monolayers (arXiv:1610.04254 [physics.bio-ph])
- [48] Krajnc M, Dasgupta S, Zihler P Ž and Prost J 2018 Fluidization of epithelial sheets by active cell rearrangements *Phys. Rev. E* **98** 022409
- [49] Sussman D M, Paoluzzi M, Cristina Marchetti M and Lisa Manning M 2018 Anomalous glassy dynamics in simple models of dense biological tissue *Europhys. Lett.* **121** 36001
- [50] Aliee M 2013 Dynamics and mechanics of compartment boundaries in developing tissues *PhD Thesis* Technische Universität Dresden
- [51] Baret J-C, Vandembroucq D and Roux S 2002 Extremal model for amorphous media plasticity *Phys. Rev. Lett.* **89** 195506
- [52] Picard G, Armand A, Lequeux F and Bocquet L 2005 Slow flows of yield stress fluids: complex spatiotemporal behavior within a simple elastoplastic model *Phys. Rev. E* **71** 010501
- [53] Kapteijns G, Ji W, Brito C, Wyart M and Lerner E 2019 Fast generation of ultrastable computer glasses by minimization of an augmented potential energy *Phys. Rev. E* **99** 012106

- [54] Kim S, Wang Y and Hilgenfeldt S 2018 Universal features of metastable state energies in cellular matter *Phys. Rev. Lett.* **120** 248001
- [55] Kirkpatrick T R and Thirumalai D 1988 Comparison between dynamical theories and metastable states in regular and glassy mean-field spin models with underlying first-order-like phase transitions *Phys. Rev. A* **37** 4439–48
- [56] Whitelam S, Berthier L and Garrahan J P 2004 Dynamic criticality in glass-forming liquids *Phys. Rev. Lett.* **92** 185705
- [57] Martens K, Bocquet L and Barrat J-L 2011 Connecting diffusion and dynamical heterogeneities in actively deformed amorphous systems *Phys. Rev. Lett.* **106** 156001
- [58] Nicolas A, Martens K, Bocquet L and Barrat J-L 2014 Universal and non-universal features in coarse-grained models of flow in disordered solids *Soft Matter* **10** 4648–61
- [59] Tyukodi B, Patinet S, Roux S and Vandembroucq D 2016 From depinning transition to plastic yielding of amorphous media: a soft-modes perspective *Phys. Rev. E* **93** 063005
- [60] Dye N A, Popović M, Spann S, Etournay R, Kainmüller D, Ghosh S, Myers E W, Jülicher F and Eaton S 2017 Cell dynamics underlying oriented growth of the drosophila wing imaginal disc *Development* **144** 4406–21
- [61] Etournay R, Merkel M, Popović M, Brandl H, Dye N A, Benoit A, Salbreux G, Eaton S and Frank J 2016 Tissueminer: a multiscale analysis toolkit to quantify how cellular processes create tissue dynamics *eLife* **5** e14334
- [62] Waddington C H 1940 The genetic control of wing development in drosophila *J. Genet.* **41** 75–113
- [63] Furuse M, Izumi Y, Oda Y, Higashi T and Iwamoto N 2014 Molecular organization of tricellular tight junctions *Tissue Barriers* **2** e28960
- [64] Bosveld F, Wang Z and Bellaïche Y 2018 Tricellular junctions: a hot corner of epithelial biology *Curr. Opin. Cell Biol.* **54** 80–8
- [65] Maloney C E and Lemaitre A 2006 Amorphous systems in athermal, quasistatic shear *Phys. Rev. E* **74** 016118
- [66] Merkel M, Etournay R, Popović M, Salbreux G, Eaton S and Frank J 2017 Triangles bridge the scales: quantifying cellular contributions to tissue deformation *Phys. Rev. E* **95**
- [67] Blanchard G B, Kabla A J, Schultz N L, Butler L C, Sanson B, Gorfinkiel N, Mahadevan L and Adams R J 2009 Tissue tectonics: morphogenetic strain rates, cell shape change and intercalation *Nat. Methods* **6** 458–64
- [68] Guirao B, Rigaud S U, Bosveld F, Bailles A, López-Gay J, Ishihara S, Sugimura K, Graner F and Bellaïche Y 2015 Unified quantitative characterization of epithelial tissue development *eLife* **4** e08519
- [69] Harris A R, Peter L, Bellis J, Baum B, Kabla A J and Charras G T 2012 Characterizing the mechanics of cultured cell monolayers *Proc. Natl Acad. Sci.* **109** 16449–54
- [70] Chattoraj J, Caroli C and Lemaitre A 2010 Universal additive effect of temperature on the rheology of amorphous solids *Phys. Rev. Lett.* **105** 266001